# CASE REPORT



# A rare case of a double high-risk gastrointestinal stromal tumor of jejunum with KIT-negative/PDGFRA-positive immunophenotype

MĂDĂLINA BOŞOTEANU<sup>1,2)</sup>, GABRIELA IZABELA BĂLŢĂTESCU<sup>1,3)</sup>, MARIANA DEACU<sup>1,2)</sup>, MARIANA ASCHIE<sup>1,2)</sup>, CĂTĂLIN ADRIAN BOSOTEANU<sup>1)</sup>

#### **Abstract**

Gastrointestinal stromal tumors (GISTs) are mesenchymal tumors that represent the second most common type in the gastrointestinal system, but clinical outcomes vary due to complex molecular changes. The aim of our study is to highlight a unique case of a 5th decade male, presenting a double primary GIST of the jejunum, associated with a rare immunophenotype and with a high risk of malignancy. Ancillary studies were performed using several biomarkers [cluster of differentiation (CD) 117/c-kit, discovered on GIST1 (DOG1), desmin, S-100, vimentin, cytokeratin 7 (CK7), Ki67, actin, platelet-derived growth factor receptor alpha (PDGFRA) and CD34], in order to confirm the diagnosis and to evaluate prognostic and predictive factors. A KIT-negative/PDGFRA-positive immunophenotype was obtained in our case and it was associated with a poor prognosis. Its unfavorable clinical evolution was sustained by recurrence as malignant (GIST) with dedifferentiation and metastases developed in less than one year after the initial diagnosis. Clinico-morphological features of GISTs with an impact on survival must be identified and a tailored therapy should be applied for each case.

**Keywords:** gastrointestinal tumor, jejunum, immunohistochemistry, dedifferentiation.

#### ☐ Introduction

Gastrointestinal stromal tumors (GISTs) are mesenchymal tumors that represent the second most common type in the gastrointestinal (GI) system after adenocarcinomas [1]. It was proved that they originate from the interstitial cells of Cajal (ICC) and have a similar immunohistochemical profile: both express the cluster of differentiation (CD) 34 and KIT mutation [2]. GISTs can be identified in any portion of the GI tract and the small bowel location occupies the second place, with a 31.8% recorded frequency after gastric involvement, which is encountered in 55.6% of cases [1]. The vast majority of cases located in the small bowel are identified in ileum and jejunum, and only 5% in the duodenum [3]. Median age is 60 years old, but any age between 10 and 100 years old can be affected with an almost equal distribution between female and male gender [4].

Even if GISTs share similar morphological features, the clinical outcomes vary, depending on the either benign or malignant characters of the proliferation that diverge due to complex molecular changes. The oncogenic *KIT* mutations are the most frequent encountered in GIST, with 80–95% incidence recorded [5], mostly in exon 11 and rarely in exons 9 and 13, which can be usually highlighted by immunohistochemistry (IHC) [6]. Another mutation, which can be associated with GIST, is that one which involved platelet-derived growth factor receptor (*PDGFR*) gene, present in 6.5% of all cases [5, 7]. All these discoveries associated with tyrosine kinase inhibitor (TKI)

therapy (Imatinib or Trastuzumab) had a great impact on survival of patients [8]. However, a small percentage of cases had a poor clinical course it was proved that these lack *KIT* or *PDGFR* mutations and there were designated as wild-type GIST (WT-GIST), with a poor prognosis [9].

The aim of our study is to highlight the unique case of a 5<sup>th</sup> decade male, presenting a double primary GIST of the jejunum. Notably, this tumor is associated with a rare immunophenotype and with a high risk of malignancy, proved by an unfavorable clinical evolution, fact sustained by the recurrence and metastases developed in less than one year after the initial diagnosis.

## ☐ Case presentation

We report a rare case of a double small bowel GIST in a 42-year-old obese and chronic smoking male patient. His past medical history revealed a left pulmonary tumor in May 2016. The tumor was large, measuring 15×10 cm, and it was located in the superior lobe, with extension to the inferior lobe. The left lung along with regional mediastinal lymph node resection have been performed in other medical center and the pathological report of the surgical specimen proved to be a pleomorphic carcinoma with metastasis in one mediastinal lymph node. Adjuvant chemotherapy was applied after surgery and follow-up in the next months noted a good clinical evolution. In June 2017, he was referred to the Department of Emergency, "Sf. Apostol Andrei" Emergency County Hospital, Constanța, Romania, accusing abdominal pain and vomiting. Abdominal

<sup>1)</sup> Clinical Service of Pathology, "Sf. Apostol Andrei" Emergency County Hospital, Constanța, Romania

<sup>&</sup>lt;sup>2)</sup>Department of Pathology, Faculty of Medicine, "Ovidius" University of Constanţa, Romania

<sup>&</sup>lt;sup>3)</sup>Center for Research and Development of the Morphological and Genetic Studies of Malignant Pathology, "Ovidius" University of Constanţa, Romania

distension and tenderness were noticed at the physical examination. The ultrasound evaluation showed dilated, fluid-filled loops of the small bowel and an area of narrowed lumen, suggestive for intestinal occlusion. Laboratory tests demonstrated a severe iron-deficiency anemia. An enterectomy was performed and the surgical

specimen was sent to the Department of Pathology. Macroscopically, two lesions were identified in the 32 cm of jejunum, with free resection margins: the first, slightly elevated, measuring 2.2 cm and the second one, a pedunculated mass of 4.5 cm in greatest dimension (Figure 1).

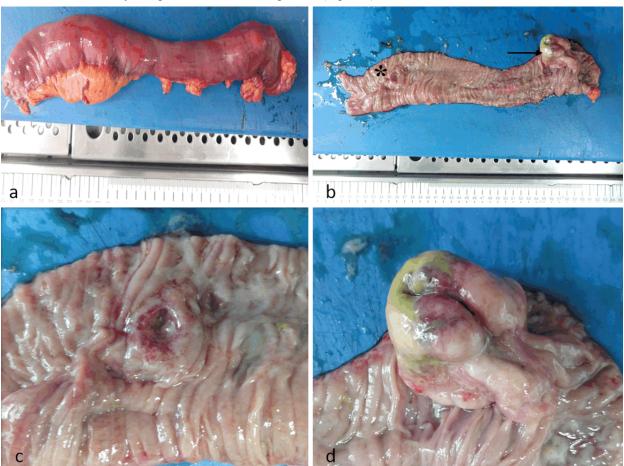


Figure 1 – Macroscopic aspect of enterectomy specimen: (a) Serosa has no macroscopically modification; (b) Cut section of the surgical specimen revealed a double tumor; (c) The first lesion is slightly elevated and ulcerated with 2.2 cm in greatest diameter (\*); (d) The second tumor is a pedunculated lesion with 4.5 cm in greatest dimension and covered with necrotic deposits on the surface.

The morphological features of these double tumors were similar and strongly suggestive for GIST, with a high risk of malignancy, being hypercellular neoplasms, predominantly composed of spindle cells with focal perinuclear vacuolization, arranged in short fascicles that randomly intersect. An epithelioid component represented by polygonal cells with eosinophilic or clear cytoplasm, arranged in nests or sheets, was accompanying the previously described population. The nuclear pleomorphism was moderate, but with more than 5 mitoses/50 high-power fields (HPFs). The neoplastic cells infiltrated the lamina propria of the mucosa, submucosa, muscular layer and subserosa (Figure 2). Tumoral necrosis, focal hemorrhage, angiotropism were morphological features also present in both lesions.

Immunohistochemical tests were performed on 4  $\mu$ m-thick sections of formalin-fixed, paraffin-embedded tissue blocks of the tumor. After the epitope retrieval, tissue sections were incubated with the following antibodies from Biocare Medical (ready-to-use): CD117/c-kit (EP100 clone), discovered on GIST1 (DOG1) (DOG1.1 clone),

desmin (D-33 clone), S-100 (15E2E2+4C4.3 clone), vimentin (SP20 clone), cytokeratin 7 (CK7) (BC-1 clone) and Ki67 (SP6 clone). We used 3,3'-diaminobenzidine (DAB) as chromogen, with brown staining. The sections were finally counterstained with Mayer's Hematoxylin. A strong nuclear and diffuse immunostaining was obtained for Ki67, with an 80% proliferative index (Figure 3). The IHC reaction for vimentin was also diffusely and strongly positive in the cytoplasm of tumor cells (Figure 3). A negative reaction was noticed in the rest of the antibodies (Figure 4). The absence of a positive immunostaining for CD117 or DOG1 raised the suspicion of a WT-GIST phenotype and further immunohistochemical tests were required. Moreover, actin, PDGFR alpha (PDGFRA) and CD34 immunostainings were subsequently evaluated in another medical unit. A negative reaction of tumor cells for actin and a positive one for the last two confirmed the initial diagnosis.

A year later, in July 2018, the tumor relapsed and an abdominal computed tomography (CT) scan identified both a tumoral lesion in the GI tract and a liver metastasis. Therefore, a second enterectomy was performed and

revealed a tumoral block due to an infiltrating and ulcerated 9 cm lesion, adherent to adjacent loops (Figure 5). The morphological evaluation emphasized a highly cellular tumor with predominantly epithelioid malignant cells and only focal spindle-shape cells, with more than 5 mitotic figures/50 HPFs and focal multinucleated giant tumor cells

(Figure 5). The microscopic features were consistent with a recurrent malignant GIST with dedifferentiation. No metastasis was identified in the 45 regional lymph nodes. Unfortunately, the prognosis was poor and the patient died shortly after surgery. All these data were collected with the written approval of the patient.

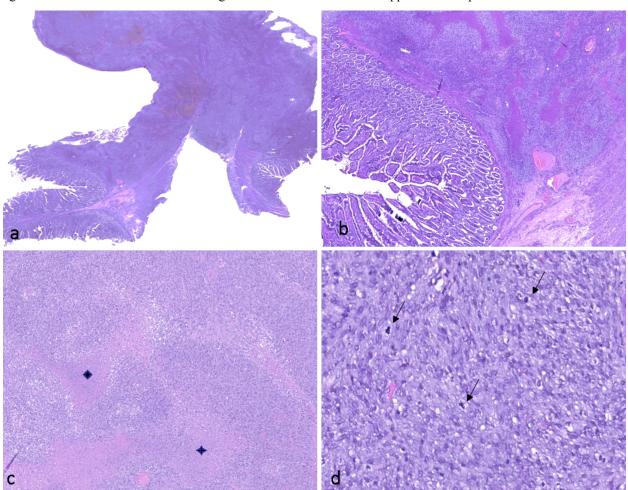


Figure 2 – Morphological features of the tumor: (a) A malignant proliferation of spindle-type cells arranged in fascicles; (b) The tumoral cells infiltrate submucosa and lamina propria of the mucosa; (c) Areas of necrosis are identified inside the tumor mass ( $\star$ ); (d) On higher objective, numerous mitoses were counted. Hematoxylin–Eosin (HE) staining: (a) Scanned image; (b and c)  $\times$ 40; (d)  $\times$ 100.

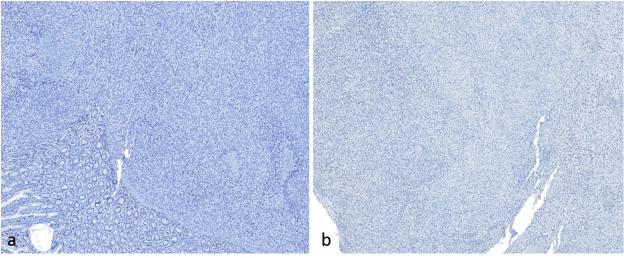


Figure 3 – Immunohistochemical examination: (a) Negative immunostaining for CD117 biomarker; (b) Negative reaction for DOG1 biomarker. Immunomarking for: (a) Anti-CD117 antibody, ×40; (b) Anti-DOG1 antibody, ×40. CD117: Cluster of differentiation 117; DOG1: Discovered on GIST1; GIST: Gastrointestinal stromal tumor.

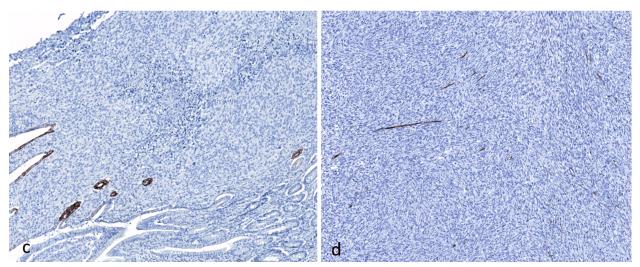


Figure 3 (continued) – Immunohistochemical examination: (c) No positive reaction for CK7 biomarker; (d) Negative reaction for desmin biomarker. Immunomarking for: (c) Anti-CK7 antibody, ×40; (d) Anti-desmin antibody, ×40. CK7: Cytokeratin 7.

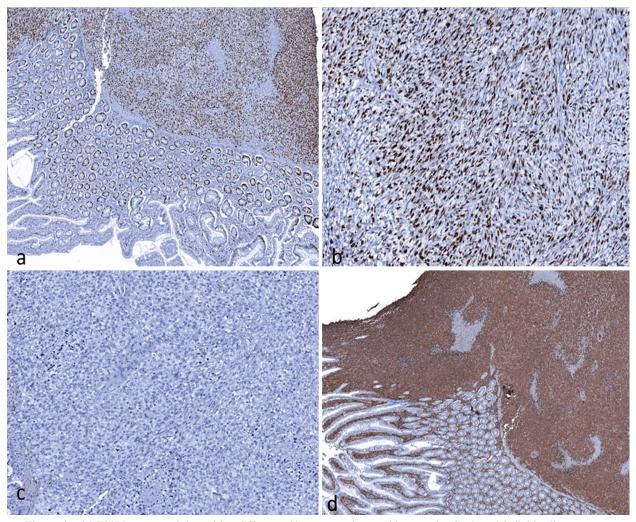


Figure 4 – (a) Ki67 immunostaining with a diffuse and intense nuclear positive reaction in the epithelioid cell component of tumor cells; (b) Similar immunohistochemical reaction for Ki67 biomarker in the spindle cell component of tumor cells; (c) Negative immunostaining for S-100 biomarker; (d) Intense and diffuse response of malignant cells to vimentin biomarker. Immunomarking for: (a and b) Anti-Ki67 antibody,  $\times$ 40 and  $\times$ 100, respectively; (c) Anti-S-100 antibody,  $\times$ 40; (d) Anti-vimentin antibody,  $\times$ 40.

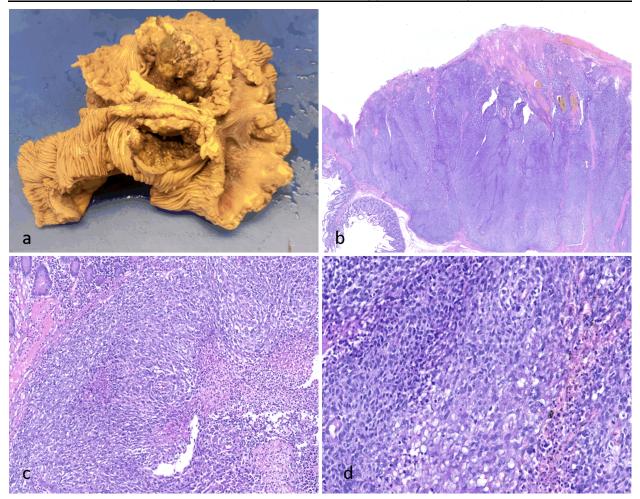


Figure 5 – (a) Macroscopic aspect of recurrence tumor with an adherently tumoral bloc of the enterectomy specimen, due to an ulcerative and infiltrative tumor; (b) Morphological features of recurrent malignant GIST with invasion of all intestinal layer; (c) There are an extensive epithelioid component and numerous areas of tumoral necrosis; (d) The content of tumor proliferation stands frequent atypical mitosis and nuclear pleomorphism. HE staining: (b) Scanned image; (c)  $\times 40$ ; (d)  $\times 100$ . GIST: Gastrointestinal stromal tumor; HE: Hematoxylin–Eosin.

#### → Discussions

GISTs are uncommon tumors and they are characterized by a wide spectrum of clinical behavior, from benign to malignant, which are caused by sequential genetic mutations. They can affect any part of the GI tract, the primary gastric location being the most frequent [1]. Small bowel involvement is rare, correspondingly the jejunum being affected in 0.1-3% of all GISTs [10]. Not only it is a rare location, but it is also associated with a poor prognosis compared to the gastric localization of a GIST, even if the morphological features are similar [4, 11]. From its first recognition, it is compulsory that factors with a predictive and prognostic value are identified and used to stratify the risk of malignancy. Several classifications have been proposed during the years, the first one being that of the National Institutes of Health (NIH) based on the tumor size and mitotic count in order to evaluate the risk for malignancy [12]. Based on these criteria, there are identified four grades for malignant risk, the most frequent were proved to be cases with a high risk (33%), followed by those with low risk (30%) [12]. In the present case, we established a high potential risk, based on the recent criterion introduced by the Armed Forces Institute of Pathology (AFIP) in the

classification, which supplementary assesses the tumor location (gastric versus other) as an additional prognostic factor [13]. In the last edition of the International Union Against Cancer (UICC) utilizing the Tumor, Node, Metastasis (TNM) Classification, the grade of a GIST was quantified, based on mitotic count, as low mitotic rate (5 or fewer mitoses/50 HPFs) and high mitotic rate (over 5 mitoses/50 HPFs), highlighting the importance of using the 40× objective (total area 5 mm<sup>2</sup> in 50 HPFs) [14]. Recent studies have shown that other morphological features may also play an important role in stratifying the prognosis. Tumoral necrosis, high cellularity, invasion of the serosa or of the adjacent structures and rich vascularity are linked to a higher risk of malignancy [15]. It was also noticed that some factors are associated with a higher risk of recurrence, like incomplete resection margin, tumor rupture and spillage during surgery [15]. In our study, we report a unique case of two primary GIST tumors located in the jejunum, penetrating the mucosa, but without reaching to the serosa and lacking perforation. Its worrisome morphological features led to frame these tumors in high-risk category for malignancy.

A tumor with morphological features suggestive for GIST must be further supplementary investigated by IHC.

Since ICCs were proved to be the original cell for GIST, the first antibody used in IHC was CD117/c-kit and its positivity is considered to be the gold standard of a precise diagnosis [13]. The KIT gene mutations, discovered by Hirota et al., are the most frequent and some of the first mutations which lead to the progression of GIST [16]. The activation of its proto-oncogene -KIT, known as CD117 (c-KIT), a receptor tyrosine kinase –, determines activation of pro-growth signals within the cells and it can be identified in up to 80–95% of GISTs [5]. Thus, it opens the way for a successful targeted therapy with Imatinib, a TKI [17]. More than that, this large group of GIST is not homogeneous in terms of prognosis and predictive factors, because an effective TKI therapy depends on the specific mutation [8]. The mutation on exon 11 (juxtamembrane domain) represents the most frequent type of KIT mutation with up to 70% of cases, the second one being those on exon 9 (extracellular dimerization) with 10–15% rates [15].

There are cases with CD117 negative and, as an alternative biomarker, DOG1/GIST1 was added, which is a transmembrane protein encoded by transmembrane member 16A (TMEM16A) gene located on chromosome 11q13 and its main role is to control the cholinergic activity of the GI smooth muscle [18, 19]. DOG1 is involved in GIST progression, independently of the KIT and PDGFRA activation and it is positive in up to 85% of GISTs, especially in those cases with a KIT/PDGFRAnegative immunophenotype [18, 20]. Even if its specificity is not high, there are still strong recommendations that DOG1 is included in the first panel of antibodies together with others markers necessary for differential diagnosis [18]. In the present case, a negative immunostaining for both biomarkers have raised the possibility for a WT-GIST phenotype of this tumor, but it was proved that there is a positive immunohistochemical expression for PDGFRA. PDGFRA is another gene with an important role in the pathogenesis of GISTs, present in 6.5% of all cases [5, 7]. One of the most important mutations of this gene is that on the exon 18, which is associated not only with resistance to Imatinib, but also to Sunitinib, a second-line TKI [3, 7]. Several studies proved that between cases with KIT mutant GIST and those with PDGFRA mutation, there are differences not only on molecular level, but also concerning the morphological traits. GISTs cases associated with PDGFRA mutation present more frequently an epithelioid morphology [21]. Moreover, it was demonstrated that the vast majority of PDGFRA mutated GIST cases are associated with the gastric, omentum or mesentery location, with a 95% rate and are less observed in the small bowel location, which harbors more frequent mutations on KIT gene [9, 21]. A few cases of GISTs do not have KIT or PDGFRA mutations and were previously considered as WT-GIST. These tumors have a higher prevalence in younger patients and have a different clinical behavior compared to those with KIT or PDGFRA mutations [22]. Some authors recommended avoiding the WT-GIST terminology, because there are other rare gene mutations involved, like the B-Raf proto-oncogene, serine/threonine kinase (BRAF) gene mutations localized on exon 15 V600E [23] or those on the succinate dehydrogenase (SDH) gene, which encodes the protein succinate dehydrogenase and is present in 90% of pediatric GISTs, compared with 12% to 15% in adult GISTs [15, 24]. It was also recommended to mention the genes which are not involved beside the WT terminology, like *KIT/PDGFRA* WT-GIST or *KIT/PDGFRA/BRAF* WT-GIST.

Numerous evidences regarding the prognostic value of Ki67 index have been collected in the last years. The research conducted by Pyo et al. included 1967 GIST cases from 24 other scientific reports, demonstrating a strong correlation between a high Ki67 and a poor prognosis. with low disease-free and overall survival rates [25]. The meta-analysis study conducted by Zhou et al. revealed that NIH-intermediate and NIH-high GIST categories are associated with higher rates of overexpression of Ki67 than the low and very low groups [26]. Zhao et al. demonstrated that the Ki67 index can be used as a predictor factor, as it was observed that a Ki67 index higher than 8% is associated with a poor response to adjuvant Imatinib therapy [27]. It was also demonstrated that the recurrence of a GIST can be predicted by a high Ki67 index [25]. In our case, a value of 80% of the Ki67 index revealed the aggressiveness of this double GIST tumor with a rapid recurrence in the dedifferentiated form.

Differential diagnoses of GIST take in consideration the morphology of the tumoral cells. Other tumors with spindle cell morphology must be differentiated by those GISTs with a predominant spindle cell proliferation. In this category, there must be included desmoid tumors, leiomyoma/leiomyosarcoma, Schwannoma, perivascular epithelioid cell tumors (PEComas) – spindle cell type. IHC may help distinguish between these entities by incorporating the following antibodies in the first panel: S-100, desmin, alpha-smooth muscle actin ( $\alpha$ -SMA),  $\beta$ -catenin antibodies. In our report, a negative reaction for S-100 and for desmin excludes other types of tumors. When an epithelioid morphology is present in a GIST, the differential category must include tumors with a similar morphology, like adenocarcinoma, PEComa with epithelioid cell or neuroendocrine tumors. In our case, taking into consideration the past history of the patient, the main differential diagnosis was with a metastasis from a lung carcinoma, but a complete negative reaction for CK7 excluded this possibility.

The gold standard treatment for GISTs without metastases is complete surgical resection (Ro), which is associated with a very good five-year survival rate of 48–70% [28]. The latest recommendations of *European Society for Medical Oncology* (ESMO) regarding the adjuvant therapy include Imatinib and other TKI [29]. Adjuvant therapy is extremely useful for GIST cases with *KIT* exon 9 mutation, but is not effective in *PDGFRA* D842V-mutated GISTs and *SDH* expression-negative GISTs [29]. Patients who have localized tumors, but are associated with a high risk of recurrence or those with unresectable tumors or with metastases must be also treated with Imatinib. In case of no response, others drugs are recommended as second-line or as third-line treatment (Regorafenib, Sunitinib) [29].

The dedifferentiation process is rare in GISTs, even if it is common in other types of sarcoma. It was first described in 2005 as a shift from a typical morphology

of a CD117-positive GIST to unrecognizable features with CD117-negative tumor recurrence after TKI treatment [30]. An anaplastic or epithelioid morphology is frequently reported in these cases, compared to the rarer angiosarcomatous aspects [31]. In our report, knowing the past history of the patient prevented us from establishing an incorrect diagnosis, due to the unusual morphological aspect. Antonescu *et al.* proved that dedifferentiation can occur in GISTs either *de novo*, or after chronic treatment with Imatinib and it is mostly secondary to genetic instability, rather than to new additional mutations [31].

#### → Conclusions

Small bowel GISTs are rare tumors with a wide range of clinical outcomes. The assessment of the prognosis does not represent an easy task and it requires corroboration of clinical, morphological and ancillary tests, performed by a multidisciplinary team. A double small bowel GIST with a rare phenotype associated with recurrence and dedifferentiation process represent a challenge regarding therapy. Tailor therapies emerged from the last decade discoveries improved the prognosis of the patients but those with rare molecular mutations associated with worrisome morphological features usually are associated with a poor outcome. The aggressiveness of a tumor is usually associated with a lack of treatment response and death of the patient. As a further matter, the peculiar morphological traits of a recurrent GIST in the dedifferentiated form must be appraised for a veracious diagnosis.

#### **Conflict of interests**

The authors declare that they have no conflict of interests.

### Acknowledgments

This work is supported by the project ANTREPRENORDOC, in the framework of Human Resources Development Operational Programme 2014–2020, financed from the European Social Fund under the contract number 36355/23.05.2019 HRD OP/380/6/13 – SMIS Code: 123847.

This research was performed in the Center for Research and Development of the Morphological and Genetic Studies of Malignant Pathology from the "Ovidius" University of Constanța, POSCCE 2.2.1. Project ID: 1844, code SMIS: 48750, CEDMOG, contract 627/11.03.2014.

#### References

- [1] Søreide K, Sandvik OM, Søreide JA, Giljaca V, Jureckova A, Bulusu VR. Global epidemiology of gastrointestinal stromal tumours (GIST): a systematic review of population-based cohort studies. Cancer Epidemiol, 2016, 40:39–46.
- [2] Kindblom LG, Remotti HE, Aldenborg F, Meis-Kindblom JM. Gastrointestinal pacemaker cell tumor (GIPACT): gastrointestinal stromal tumors show phenotypic characteristics of the interstitial cells of Cajal. Am J Pathol, 1998, 152(5):1259– 1269.
- [3] Liegl-Atzwanger B, Fletcher JA, Fletcher CD. Gastrointestinal stromal tumors. Virchows Arch, 2010, 456(2):111–127.
- [4] Miettinen M, Sobin LH, Lasota J. Gastrointestinal stromal tumors of the stomach: a clinicopathologic, immunohistochemical, and molecular genetic study of 1765 cases with long-term follow-up. Am J Surg Pathol, 2005, 29(1):52–68.
- [5] Ricci R, Dei Tos AP, Rindi G. GISTogram: a graphic presentation of the growing GIST complexity. Virchows Arch, 2013, 463(4):481–487.

- [6] Agaram NP, Besmer P, Wong GC, Guo T, Socci ND, Maki RG, DeSantis D, Brennan MF, Singer S, DeMatteo RP, Antonescu CR. Pathologic and molecular heterogeneity in imatinib-stable or imatinib-responsive gastrointestinal stromal tumors. Clin Cancer Res, 2007, 13(1):170–181.
- [7] Heinrich MC, Corless CL, Duensing A, McGreevey L, Chen CJ, Joseph N, Singer S, Griffith DJ, Haley A, Town A, Demetri GD, Fletcher CD, Fletcher JA. *PDGFRA* activating mutations in gastrointestinal stromal tumors. Science, 2003, 299(5607): 708–710.
- [8] Corless CL, Heinrich MC. Molecular pathobiology of gastrointestinal stromal sarcomas. Annu Rev Pathol, 2008, 3:557– 586.
- [9] Lasota J, Dansonka-Mieszkowska A, Sobin LH, Miettinen M. A great majority of GISTs with PDGFRA mutations represent gastric tumors of low or no malignant potential. Lab Invest, 2004, 84(7):874–883.
- [10] Mujawar P, Nikumbh DB, Suryawanshi KH, Pagare PS, Thakur R. Malignant gastrointestinal stromal tumor (GIST) of the jejunum: the mysterious complex presentation. Int J Med Sci Public Health, 2015, 4(8):1168–1171.
- [11] Miettinen M, Makhlouf H, Sobin LH, Lasota J. Gastrointestinal stromal tumors of the jejunum and ileum: a clinicopathologic, immunohistochemical, and molecular genetic study of 906 cases before imatinib with long-term follow-up. Am J Surg Pathol, 2006, 30(4):477–489.
- [12] Fletcher CD, Berman JJ, Corless C, Gorstein F, Lasota J, Longley BJ, Miettinen M, O'Leary TJ, Remotti H, Rubin BP, Shmookler B, Sobin LH, Weiss SW. Diagnosis of gastrointestinal stromal tumors: a consensus approach. Hum Pathol, 2002, 33(5):459–465.
- [13] Miettinen M, Lasota J. Gastrointestinal stromal tumors: review on morphology, molecular pathology, prognosis, and differential diagnosis. Arch Pathol Lab Med, 2006, 130(10):1466–1478.
- [14] DeMatteo RP, Maki RG, Agulnik M, Brookland RK, Hornick JL, Jones RL, Keedy VL, Lazar AJ, O'Sullivan B, Raut CP, Tedder PS, Pollock RE. Chapter 43: Gastrointestinal stromal tumor. In: Amin MB, Edge S, Greene F, Byrd DR, Brookland RK, Washington MK, Gershenwald JE, Compton CC, Hess KR, Sullivan DC, Jessup JM, Brierley JD, Gaspar LE, Schilsky RL, Balch CM, Winchester DP, Asare EA, Madera M, Gress DM, Meyer LR (eds). AJCC Cancer Staging Manual. 8<sup>th</sup> edition, American College of Surgeons, Springer International Publishing, 2017, 523–529.
- [15] Lim KT, Tan KY. Current research and treatment for gastrointestinal stromal tumors. World J Gastroenterol, 2017, 23(27): 4856–4866
- [16] Hirota S, Isozaki K, Moriyama Y, Hashimoto K, Nishida T, Ishiguro S, Kawano K, Hanada M, Kurata A, Takeda M, Muhammad Tunio G, Matsuzawa Y, Kanakura Y, Shinomura Y, Kitamura Y. Gain-of-function mutations of *c-kit* in human gastrointestinal stromal tumors. Science, 1998, 279(5350): 577–580.
- [17] Joensuu H, Roberts PJ, Sarlomo-Rikala M, Andersson LC, Tervahartiala P, Tuveson D, Silberman S, Capdeville R, Dimitrijevic S, Druker B, Demetri GD. Effect of the tyrosine kinase inhibitor STI571 in a patient with a metastatic gastrointestinal stromal tumor. N Engl J Med, 2001, 344(14):1052– 1056.
- [18] Kang GH, Srivastava A, Kim YE, Park HJ, Park CK, Sohn TS, Kim S, Kang DY, Kim KM. DOG1 and PKC-θ are useful in the diagnosis of KIT-negative gastrointestinal stromal tumors. Mod Pathol, 2011, 24(6):866–875.
- [19] Şahin S, Ekinci Ö, Seçkin S, Dursun A. The diagnostic and prognostic utility of DOG1 expression on gastrointestinal stromal tumors. Turk Patoloji Derg, 2017, 33(1):1–8.
- [20] Baskin Y, Kocal GC, Kucukzeybek BB, Akbarpour M, Kayacik N, Sagol O, Ellidokuz H, Oztop I. PDGFRA and KIT mutation status and its association with clinicopathological properties, including DOG. Oncol Res, 2016, 24(1):41–53.
- [21] Wardelmann E, Hrychyk A, Merkelbach-Bruse S, Pauls K, Goldstein J, Hohenberger P, Losen I, Manegold C, Büttner R, Pietsch T. Association of platelet-derived growth factor receptor alpha mutations with gastric primary site and epithelioid or mixed cell morphology in gastrointestinal stromal tumors. J Mol Diagn, 2004, 6(3):197–204.

- [22] Boikos SA, Pappo AS, Killian JK, LaQuaglia MP, Weldon CB, George S, Trent JC, von Mehren M, Wright JA, Schiffman JD, Raygada M, Pacak K, Meltzer PS, Miettinen MM, Stratakis C, Janeway KA, Helman LJ. Molecular subtypes of KITIPDGFRA wild-type gastrointestinal stromal tumors: a Report from the National Institutes of Health Gastrointestinal Stromal Tumor Clinic. JAMA Oncol, 2016, 2(7):922–928.
- [23] Agaram NP, Wong GC, Guo T, Maki RG, Singer S, DeMatteo RP, Besmer P, Antonescu CR. Novel V600E BRAF mutations in imatinib-naive and imatinib-resistant gastrointestinal stromal tumors. Genes Chromosomes Cancer, 2008, 47(10):853–859.
- [24] Gaal J, Stratakis CA, Carney JA, Ball ER, Korpershoek E, Lodish MB, Levy I, Xekouki P, van Nederveen FH, den Bakker MA, O'Sullivan M, Dinjens WN, de Krijger RR. SDHB immunohistochemistry: a useful tool in the diagnosis of Carney–Stratakis and Carney triad gastrointestinal stromal tumors. Mod Pathol, 2011, 24(1):147–151.
- [25] Pyo JS, Kang G, Sohn JH. Ki-67 labeling index can be used as a prognostic marker in gastrointestinal stromal tumor: a systematic review and meta-analysis. Int J Biol Markers, 2016, 31(2):e204–e210.
- [26] Zhou Y, Hu W, Chen P, Abe M, Shi L, Tan SY, Li Y, Zong L. Ki67 is a biological marker of malignant risk of gastrointestinal stromal tumors: a systematic review and meta-analysis. Medicine (Baltimore), 2017, 96(34):e7911.
- [27] Zhao WY, Xu J, Wang M, Zhang ZZ, Tu L, Wang CJ, Lin TL, Shen YY, Liu Q, Cao H. Prognostic value of Ki67 index in gastrointestinal stromal tumors. Int J Clin Exp Pathol, 2014, 7(5):2298–2304.
- [28] Wu PC, Langerman A, Ryan CW, Hart J, Swiger S, Posner MC. Surgical treatment of gastrointestinal stromal tumors in

- the imatinib (STI-571) era. Surgery, 2003, 134(4):656-665; discussion 665-666.
- [29] Casali PG, Abecassis N, Aro HT, Bauer S, Biagini R, Bielack S, Bonvalot S, Boukovinas I, Bovee JVMG, Brodowicz T, Broto JM, Buonadonna A, De Álava E, Dei Tos AP, Del Muro XG, Dileo P, Eriksson M, Fedenko A, Ferraresi V, Ferrari A, Ferrari S, Frezza AM, Gasperoni S, Gelderblom H, Gil T, Grignani G, Gronchi A, Haas RL, Hassan B, Hohenberger P, Issels R, Joensuu H, Jones RL, Judson I, Jutte P, Kaal S, Kasper B, Kopeckova K, Krákorová DA, Le Cesne A, Lugowska I, Merimsky O, Montemurro M, Pantaleo MA, Piana R, Picci P, Piperno-Neumann S, Pousa AL, Reichardt P, Robinson MH, Rutkowski P, Safwat AA, Schöffski P, Sleijfer S, Stacchiotti S, Sundby Hall K, Unk M, Van Coevorden F, van der Graaf WTA, Whelan J, Wardelmann E, Zaikova O, Blay JY; ESMO Guidelines Committee and EURACAN. Gastrointestinal stromal tumours: ESMO-EURACAN Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol, 2018, 29(Suppl 4): iv68-iv78.
- [30] Pauwels P, Debiec-Rychter M, Stul M, De Wever I, Van Oosterom AT, Sciot R. Changing phenotype of gastrointestinal stromal tumours under imatinib mesylate treatment: a potential diagnostic pitfall. Histopathology, 2005, 47(1):41–47.
- [31] Antonescu CR, Romeo S, Zhang L, Nafa K, Hornick JL, Nielsen GP, Mino-Kenudson M, Huang HY, Mosquera JM, Dei Tos PA, Fletcher CD. Dedifferentiation in gastrointestinal stromal tumor to an anaplastic KIT-negative phenotype: a diagnostic pitfall: morphologic and molecular characterization of 8 cases occurring either de novo or after imatinib therapy. Am J Surg Pathol, 2013, 37(3):385–392.

#### Corresponding author

Gabriela Izabela Bălţătescu, MD, PhD, Clinical Service of Pathology, "Sf. Apostol Andrei" Emergency County Hospital, Constanţa; Center for Research and Development of the Morphological and Genetic Studies of Malignant Pathology, "Ovidius" University of Constanţa, 1 Universităţii Lane, 900470 Constanţa, Romania; Phone +40241–605 002, e-mail: gabrielabaltatescu@yahoo.com

Received: April 17, 2019

Accepted: December 3, 2019