ORIGINAL PAPER



Prognostic significance of cell-adhesion molecules in histological variants of papillary thyroid carcinoma

ELENA CORINA ANDRIESCU^{1,2)}, IRINA-DRAGA CĂRUNTU¹⁾, SIMONA ELIZA GIUŞCĂ¹⁾, LUDMILA LOZNEANU^{1,2)}, DELIA GABRIELA CIOBANU APOSTOL^{1,2)}

Abstract

The molecular structure of E-cadherin and its function are intimately related to β -catenin, their interactions ensuring the cell morphology and stability. Alterations of E-cadherin- β -catenin complex facilitate the tumor growth and spreading in the carcinogenic mechanism. We aimed to assess the E-cadherin and β -catenin immunoexpressions in different variants of papillary thyroid carcinoma (PTC), and the relationship of these markers with the clinicopathological prognostic factors. Our study group included 70 cases of PTC divided into two risk groups. The low-risk group comprised 45 cases diagnosed as conventional, follicular, oncocytic, macrofollicular, and clear cell variants, whereas the high-risk group consisted of 25 cases diagnosed as tall cell, follicular angioinvasive, cribriform-morular, hobnail, diffuse sclerosing, and solid subtype, respectively. Immunohistochemical exam was performed by using anti-E-cadherin and anti- β -catenin antibodies, and their expressions were semi-quantitatively evaluated. The association between E-cadherin and β -catenin, respectively, and clinicopathological prognostic factors was statistically analyzed. We noted statistically significant differences between membranous E-cadherin expression (low *versus* high) and tumor size, histological risk groups, tumor stage, lymph node metastases, vascular invasion and tumor relapse. We also found statistically significant correlation between membranous β -catenin expression (low *versus* high) and the risk groups, tumor size and tumor stage, but no associations of cytoplasmic β -catenin (low *versus* high) with the clinicopathological characteristics. Our study demonstrates that E-cadherin and β -catenin expressions differ in low- and high-risk groups of PTC. The aggressive behavior of the high-risk histological variants is associated with reduced membranous E-cadherin, and loss of membranous β -catenin followed by enhanced cytoplasmic expression. These results open large standpoints for a deeper characterization of the hi

Keywords: papillary thyroid carcinoma, histological variants, E-cadherin, β-catenin.

□ Introduction

Papillary thyroid carcinoma (PTC) is the widespread type of thyroid carcinoma (TC) and represents 70-85% of this malignancy with an increased global incidence during the last years [1]. PTC is usually correlated with a good prognostic, but certain patients can have an aggressive behavior of disease, due to the occurrence of local or distant metastases; consequently, a worse prognostic of PTC cannot be excluded [2]. Although a solid panel of classical, clinicopathological prognostic factors is already defined [3], it seems that they are not enough to explain the variability of PTC progression. Therefore, the understanding of these different courses of PTC is far to be completed. As a consequence, the interest in carcinogenesis mechanism focuses on the study of new prognostic factors that could clarify the distinctive comportment of the tumor cells within the thyroid environment [4–7].

One challenging research topic address the molecular markers that include molecules involved in the cellular cycle regulation (*i.e.*, cyclin D1, p27, p53), or cell growth (Her2/neu), in cell adhesion (E-cadherin, β -catenin, claudin-1, tubulin), or in tumor microenvironment changes (fibronectin) [4, 8, 9]. Concurrently, the histological variants of PTC [1] sustain the stratification of PTC cases in high-risk or low-risk categories, starting from the time of diagnosis [10–14]. In spite of the strong attempt to

validate new proposed prognostic factors, the diversity of pro and cons arguments sustains the actuality of this research.

E-cadherin, a calcium-dependent transmembrane cell adhesion molecule, is essential for the adhesion and normal function of epithelial cells [15]. In the adherens junctions, the E-cadherin intracytoplasmic domain links to β -catenin that in turn connects to α -catenin jointed to the actinic cytoskeleton [16, 17]. Besides the role in the intercellular stability [18], β -catenin acts as a signaling factor in the canonical Wnt pathway [19]. The decrease of E-cadherin expression is responsible for the loss of cell adhesion, tumor growth and proliferation, leading to metastasis [20, 21]. Low E-cadherin expression has been reported in several malignancies, in association with tumor advanced stages and disease progression [22–25]. Several studies sustain that loss of E-cadherin is a decisive step in dedifferentiation, progression, and metastatic spread of TC [26–28]. Thus, E-cadherin expression is maintained in differentiated or in minimally invasive TC, and completely absent in undifferentiated ones. On the other hand, the involvement of β -catenin in carcinogenesis is also documented in different types of tumors [29-33], but few reports focus on its expression in TC, in relationship with a poor prognostic [34–40].

Starting from this point, the aim of our study was to analyze the expression pattern of E-cadherin and β -catenin

¹⁾Department of Morphofunctional Sciences I, "Grigore T. Popa" University of Medicine and Pharmacy, Iaşi, Romania

²⁾Department of Pathology, "Sf. Spiridon" Emergency County Hospital, Iaşi, Romania

in different subtypes of PTC and the relationship of these markers with the clinicopathological factors.

Our study group included 70 cases of PTC, diagnosed between 2006 and 2016 at the Laboratory of Pathology, "Sf. Spiridon" Emergency County Hospital, Iaşi, Romania. All cases were histopathologically reassessed by three pathologists for ascertain the PTC histological subtype and then divided into two risk groups [10–14]. The low-risk group comprised 45 cases diagnosed as conventional, follicular, oncocytic, macrofollicular, and clear cell variants, whereas the high-risk group consisted of 25 cases diagnosed as tall cell, follicular angioinvasive, cribriform-morular, hobnail, diffuse sclerosing, and solid subtype, respectively.

The study has been approved by the Ethics Committee of the "Grigore T. Popa" University of Medicine and Pharmacy, Iaşi, based on the patients' informed consent.

Immunohistochemistry (IHC)

The sections obtained from the selected paraffinembedded blocks corresponding to these cases were dewaxed in xylene, rehydrated in consecutive descending concentrations of ethanol (100%, 90%, 80%, and 70%), and rinsed in distillated water. The antigen retrieval was made by using Heat-Induced Epitope Retrieval (HIER) procedure, with an antigen retrieval solution with pH 6, in a microwave oven, for 30 minutes. After the endogenous peroxidase blocking with 3% hydrogen peroxide, the sections were incubated with the primary antibodies E-cadherin (clone EP700Y, 1:100 dilution, Thermo Scientific, USA) and β -catenin (clone β -catenin-1, 1:300 dilution, Agilent-Dako, USA). The reaction was amplified with UltraVision Quanto Detection System Horseradish peroxidase (HRP) DAB (Thermo Scientific, USA) and developed with 3.3'-diaminobenzidine (DAB) tetrahydrochloride chromogen (DakoCytomation, Carpinteria, USA). The counterstaining of the sections was done with Mayer's Hematoxylin. The normal thyroid tissue or the benign associated pathology were used as internal positive controls for the two antibodies.

Semi-quantitative assessment

The semi-quantitative assessment was done by using adapted scores based on the literature reports [36, 41] that took into account the staining intensity (I) and the percentage of positive cells (P).

For E-cadherin, we assessed the membranous expression, whereas the β -catenin expression was quantified both at membranous and cytoplasmic level. The intensity of the immunoreaction for the two markers was scored as: 0 – absent, 1 – weak, 2 – moderate, and 3 – strong. The percentage of E-cadherin membranous positive cells was scored as follows: 0 – <5%, 1 – 6–25%, 2 – 26–50%, 3 – 51–77%, and 4 – >75%. The E-cadherin final score was obtained by multiplying P by I. E-cadherin score values between 1–4 were considered low, and score values between 6–12 were considered high. The percentage of β -catenin membranous or cytoplasmic immunopositive

cells was scored as: 0 - <10%, 1 - 10 - 30%, 2 - 31 - 50%, 3 - 51 - 70%, 4 - >70%. The β -catenin final score resulted by summation of P and I. Cases with values between 1 - 3 were considered with low score, and cases with score between 4 - 7 were considered with high score.

Statistical analysis

Statistical analysis was performed by using Statistical Package for the Social Sciences (SPSS) ν . 19 program (SPSS Inc., IBM Corporation, Chicago, IL, USA) and the χ^2 (chi-square) test (Maximum-Likelihood, Yates, Mantel–Haenszel). Statistical significance was considered for p<0.05.

Clinicopathological characteristics

In the whole group, 55 patients were females (more than two-thirds), and 15 patients were males. At the time of the diagnosis, the age of the patients ranged between 17 and 79 years old, with a median age of 49 years old. Surgical treatment consisted in total thyroidectomy with lymphadenectomy for 41 patients, and partial thyroidectomy for the remaining 29 cases. The histopathological exam revealed the tumor extrathyroidal extension in 49 cases, presence of lympho-vascular invasion in 46 cases, and multifocal tumors in 25 cases. The distribution of the PTC histological variants was as follows: conventional subtype – 16 cases, follicular subtype – nine cases, macrofollicular - six cases, clear cell - four cases, oncocytic variant – 10 cases, tall cell – eight cases, cribriform – five cases, hobnail – one case, diffuse sclerosing – three cases, solid – five cases, angioinvasive follicular – one case, conventional with dedifferentiation to squamous cell carcinoma - one case, oncocytic with undifferentiated solid areas - one case. Based on the Classification of Malignant Tumors (TNM), 18 cases were staged as pT1, 27 cases as pT2, and 25 cases as pT3. In nine patients, lymph node relapse was registered.

E-cadherin and β -catenin assessment

The intensity of E-cadherin membranous immuno-expression was predominantly moderate or low in PTC, compared to the adjacent benign or normal thyroid tissue, strongly positive. Moderate membranous expression was observed not only in conventional and follicular subtypes, but also in some cases with a more aggressive course, like tall cell, cribriform and diffuse sclerosing variants. Low E-cadherin expression was noticed in PTC cases with high tumor extent (pT3) and lymph node metastases. In these cases, E-cadherin expression was heterogeneous, with 20–60% positive cells present in tumor areas.

 β -Catenin immunostaining pattern in normal thyroid tissue or in the adjacent Hashimoto thyroiditis and nodular goiter was strong membranous, either circumferential in the cuboidal and columnar cells, either basal in the hypofunctional areas. In PTC, β -catenin had a discontinuous membranous expression with the persistence of lateral membrane staining and the absence of immunostaining in apical and basal pole of the tumor cells. We also noted that in areas where the membranous staining was lacking, β -catenin was expressed in the cytoplasm of the tumor

cells with moderate or even high intensity. Some cases of conventional PTC preserved a strong, diffuse membranous staining, whereas in the follicular, tall cell and solid variants a large heterogeneity was observed. The nuclear staining of β -catenin was identified in PTC cribriform-

morular variant in approximately 40% of the tumor cells together with moderate cytoplasmic expression and loss of the membranous one in more than 80% of the cells.

Figures 1–6 illustrate different immunoexpression patterns of E-cadherin and β -catenin.

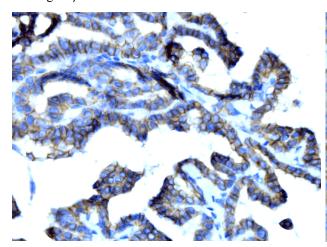


Figure 1 – PTC, conventional variant: membranous E-cadherin expression with strong intensity. IHC staining, anti-E-cadherin antibody, ×400.

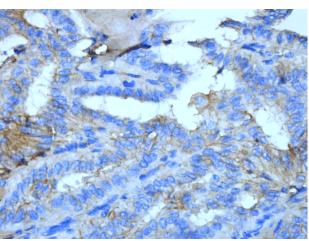


Figure 2 – PTC, tall cell variant: membranous E-cadherin expression with low intensity and heterogeneous pattern. IHC staining, anti-E-cadherin antibody, ×400.

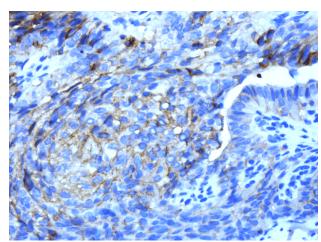


Figure 3 – PTC, cribriform-morular variant: membranous E-cadherin expression with low intensity and heterogeneous pattern. IHC staining, anti-E-cadherin antibody, ×400.

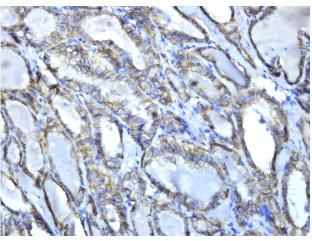


Figure 4 – PTC, follicular variant: membranous β -catenin expression with moderate intensity and homogenous pattern. IHC staining, anti- β -catenin antibody, $\times 100$.

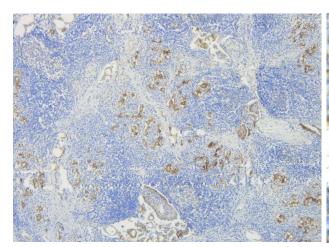


Figure 5 – PTC, diffuse sclerosing variant: cytoplasmic β -catenin expression with high intensity. IHC staining, anti- β -catenin antibody, $\times 50$.

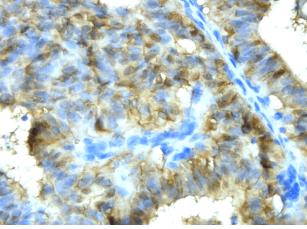


Figure 6 – PTC, cribriform-morular variant: cytoplasmic β -catenin expression with moderate intensity, associated with isolate nuclear β -catenin immunostaining. IHC staining, anti- β -catenin antibody, $\times 400$.

The results of the semi-quantitative assessment of E-cadherin and β -catenin expressions in accordance with the low and high score values, respectively, and the main clinicopathological characteristics were summarized in Tables 1 and 2.

The statistical analysis of E-cadherin expression and the clinicopathological factors showed significant differences between low *versus* high expression and tumor size (p=0.017), risk groups (p=0.003), tumor stage (p=0), lymph node metastases (p=0.001), vascular invasion (p=0) and tumor relapse (p=0.005). No correlation of E-cadherin expression with age, gender, multifocality and extrathyroidal invasion was found (Table 1). Membranous β -catenin expression (low *versus* high) was statistically significant correlated with the risk groups, tumor size and tumor stage (Table 2). No statistically significant differences were found between cytoplasmic β -catenin (low *versus* high) and the clinicopathological characteristics (Table 2).

₽ Discussions

Even though the prognostic assessment of PTC relies, according to the *World Health Organization* (WHO), on the standard clinicomorphological factors, nowadays the pathologists look on the "candidate" prognosis markers and try to validate more sensitive criteria for the assessment of neoplastic evolution. These markers could be related to the carcinogenesis mechanism, their distinctive involvement controlling the subtle differences in prognostic. The endeavor of this approach could be the identification within the same histological phenotype of new subclasses of diagnosis (at present just predicted in an intuitive way, in relationship with the tumor behavior), characterized by molecular features.

Based on the expertise of the "Sf. Spiridon" Emergency County Hospital of Iaşi in the diagnosis and surgical treatment of thyroid tumor pathology, in the last years our scientific interest was pointed on the identification of a stratification algorithm applicable in PTC [40, 42–44]. This item comes out from the molecular classification of

breast cancer [45] and, more recent, of lung cancer [46], that changed the traditional concept of diagnosis and treatment.

Table 1 – Relationship between E-cadherin expression and clinicopathological characteristics

characteristics Low (n=42) High (n=28) square test Age at diagnosis <55 years old 15 (35.7%) 27 (64.3%) 1 ≥55 years old 10 (35.7%) 18 (64.3%) 1 Gender Female 20 (36.4%) 35 (63.6%) 0.828 Male 5 (33.3%) 10 (66.7%) 1 Tumor size (median) <30 mm 6 (20%) 24 (80%) 0.017 ≥30 mm 19 (47.5%) 21 (52.5%) 0.017 Histopathological subtype Low-risk group 11 (22.7%) 34 (77.3%) 0.003 High-risk group 15 (57.7%) 10 (42.3%) 0.003 Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.577	Clinicopathological	E-cadherin n	p-value from chi-						
<55 years old 15 (35.7%) 27 (64.3%) 1 ≥55 years old 10 (35.7%) 18 (64.3%) 1 Gender Female 20 (36.4%) 35 (63.6%) 0.828 Male 5 (33.3%) 10 (66.7%) 0.828 Tumor size (median) <30 mm 6 (20%) 24 (80%) 0.017 ≥30 mm 19 (47.5%) 21 (52.5%) 0.017 Histopathological subtype Low-risk group 11 (22.7%) 34 (77.3%) 0.003 High-risk group 15 (57.7%) 10 (42.3%) 0.003 Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.577		Low	High	•					
Segment	Age at diagnosis								
≥55 years old 10 (35.7%) 18 (64.3%) Gender Female 20 (36.4%) 35 (63.6%) 0.828 Male 5 (33.3%) 10 (66.7%) 0.828 Tumor size (median) <30 mm 6 (20%) 24 (80%) 24 (50%) 24 (52.5%) Histopathological subtype Low-risk group 11 (22.7%) 34 (77.3%) 0.003 High-risk group 15 (57.7%) 10 (42.3%) 0.003 Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.577	<55 years old	15 (35.7%)	27 (64.3%)	1					
Female 20 (36.4%) 35 (63.6%) 0.828 Male 5 (33.3%) 10 (66.7%) 0.828 Tumor size (median) <30 mm 6 (20%) 24 (80%) 0.017 Example 19 (47.5%) 21 (52.5%) 0.017 Histopathological subtype Low-risk group 11 (22.7%) 34 (77.3%) 0.003 High-risk group 15 (57.7%) 10 (42.3%) 0.003 Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.577	≥55 years old	10 (35.7%)	18 (64.3%)	- 1					
Male 5 (33.3%) 10 (66.7%) Tumor size (median) <30 mm 6 (20%) 24 (80%) ≥30 mm 19 (47.5%) 21 (52.5%) Histopathological subtype Low-risk group 11 (22.7%) 34 (77.3%) High-risk group 15 (57.7%) 10 (42.3%) Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.577									
Male 5 (33.3%) 10 (66.7%) Tumor size (median) <30 mm	Female	20 (36.4%)	35 (63.6%)	0.828					
<30 mm 6 (20%) 24 (80%) ≥30 mm 19 (47.5%) 21 (52.5%) Histopathological subtype Low-risk group 11 (22.7%) 34 (77.3%) High-risk group 15 (57.7%) 10 (42.3%) Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.577	Male	5 (33.3%)	10 (66.7%)	0.020					
230 mm 19 (47.5%) 21 (52.5%) Histopathological subtype Low-risk group 11 (22.7%) 34 (77.3%) High-risk group 15 (57.7%) 10 (42.3%) Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.017 0.003	Tumor size (median)								
≥30 mm 19 (47.5%) 21 (52.5%) Histopathological subtype Low-risk group 11 (22.7%) 34 (77.3%) High-risk group 15 (57.7%) 10 (42.3%) Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.577	<30 mm	<30 mm 6 (20%) 24 (80%)							
Low-risk group 11 (22.7%) 34 (77.3%) High-risk group 15 (57.7%) 10 (42.3%) Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.577	≥30 mm	19 (47.5%)	21 (52.5%)	0.017					
High-risk group 15 (57.7%) 10 (42.3%) Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.577	` ' ' ' ' '								
High-risk group 15 (57.7%) 10 (42.3%) Focality of the tumor Unifocal 15 (33.3%) 30 (66.7%) 0.577	Low-risk group	11 (22.7%)	34 (77.3%)	0.000					
Unifocal 15 (33.3%) 30 (66.7%) 0.577	High-risk group	15 (57.7%)	10 (42.3%)	0.003					
0.577	Unifocal	15 (33.3%)	30 (66.7%)	0.577					
Multifocal 10 (40%) 15 (60%)	Multifocal	10 (40%)	15 (60%)	0.577					
Tumor stage									
T1 + T2 8 (17.8%) 37 (82.2%)	T1 + T2	8 (17.8%)	37 (82.2%)	0					
T3 17 (68%) 8 (32%)	Т3	17 (68%)	8 (32%)						
Lymph node metastases									
NO 3 (15.8%) 16 (84.2%)	N0	3 (15.8%)	16 (84.2%)	0.004					
N1 15 (68.2%) 7 (31.8%) 0.001	N1	15 (68.2%)	7 (31.8%)	0.001					
Lympho-vascular invasion									
Absent 1 (4.2%) 23 (95.8%)	Absent 1 (4.2%) 23 (95.8%)		•						
Present 24 (52.2%) 22 (47.8%)	Present	24 (52.2%)	22 (47.8%)	- 0					
Extrathyroidal invasion									
Absent 5 (23.8%) 16 (76.2%)	Absent	5 (23.8%)	16 (76.2%)	0.174					
Present 20 (40.8%) 29 (59.2%) 0.174	Present	20 (40.8%)	29 (59.2%)	- 0.174					
Tumor relapse									
Absent 18 (29.5%) 43 (70.5%) 0.005	Absent	18 (29.5%)	43 (70.5%)	0.005					
Present 7 (77.8%) 2 (22.2%)	Present	7 (77.8%)	2 (22.2%)	0.005					

Table 2 – Relationship between β -catenin expression and clinicopathological characteristics

Clinicopathological _ characteristics	β-Catenin membranous expression		<i>p</i> -value – from <i>chi</i> - –	β-Catenin cytoplasmic expression		p-value
	Low (n=42)	High (n=28)	square test	Low (n=36)	High (n=34)	 from chi- square test
		Age	at diagnosis			
<55 years old	25 (59.5%)	17 (40.5%)	- 0.921 -	20 (47.6%)	22 (52.4%)	- 0.473
≥55 years old	17 (60.7%)	11 (39.3%)	- 0.921 -	16 (57.1%)	12 (42.9%)	- 0.473
			Gender			
Female	31 (56.4%)	24 (43.6%)	- 0.234 -	26 (47.3%)	29 (52.7%)	- 0.247
Male	11 (73.3%)	4 (26.7%)		10 (66.7%)	5 (33.3%)	
		Tumo	r size (median)			
<30 mm	16 (47.1%)	18 (52.9%)	- 0.032 -	16 (47.1%)	18 (52.9%)	- 0.633
≥30 mm	26 (72.2%)	10 (27.8%)		20 (55.6%)	16 (44.4%)	
		Histopat	hological subtype			
Low-risk group	23 (51.1%)	22 (48.9%)	0.040	26 (59.1%)	18 (40.9%)	- 0.095
High-risk group	19 (76%)	6 (24%)	– 0.042 –	10 (38.5%)	16 (61.5%)	- 0.095
		Focal	ity of the tumor			
Unifocal	26 (54.2%)	22 (45.8%)	- 0.141 -	27 (56.3%)	21 (43.7%)	- 0.305
Multifocal	16 (72.7%)	6 (27.3%)		9 (40.9%)	13 (59.1%)	

Clinicopathological _ characteristics	β-Catenin membranous expression		p-value	β-Catenin cytoplasmic expression		p-value
	Low (n=42)	High (n=28)	 from chi- square test 	Low (n=36)	High (n=34)	from chi- square test
		T	umor stage			
T1 + T2	24 (51.1%)	23 (48.9%)	- 0.029 -	24 (51.1%)	23 (48.9%)	- 0.93
T3	18 (78.3%)	5 (21.7%)		12 (52.2%)	11 (48.8%)	
		Lymph	node metastases			
N0	14 (73.7%)	5 (26.3%)	- 0.121 -	12 (63.2%)	7 (36.8%)	- 0.257
N1	11 (50%)	11 (50%)		10 (45.5%)	12 (54.5%)	
		Lympho	-vascular invasion	1		
Absent	14 (51.9%)	13 (48.1%)	0.07	15 (55.6%)	12 (44.4%)	- 0.63
Present	28 (65.1%)	15 (34.9%)	– 0.27 –	21 (48.8%)	22 (51.2%)	- 0.63
		Extratl	nyroidal invasion			
Absent	12 (57.1%)	9 (42.9%)	- 0.749 -	9 (42.9%)	12 (57.1%)	- 0.437
Present	30 (61.2%)	19 (38.8%)		27 (55.1%)	22 (44.9%)	
		Tu	mor relapse			
Absent	36 (59%)	25 (41%)	0.663	29 (47.5%)	32 (52.5%)	0.450
Present	6 (66.7%)	3 (33.3%)	- 0.662 -	7 (77.8%)	2 (22.2%)	- 0.152

Within this context, the abnormalities of adhesion molecules could offer a more comprehensive understanding of the variances in tumor behavior. The molecular structure of E-cadherin and its function are intimately related to β -catenin, their interactions ensuring the normal cell morphology and stability [16–18]. Alterations in E-cadherin lead to the increase of cytoplasmic β -catenin expression and, subsequently, to the amplification of transcription [47]. This phenomenon, regarded as a sequence of the carcinogenic mechanisms, facilitates the tumor growth and spreading.

Starting from the '90, several studies focus on E-cadherin changes in PTC and follicular thyroid carcinoma (FTC) and show a significant loss of its membranous expression in poorly and undifferentiated forms, respectively [48–55]. Conversely, in benign thyroid lesions, a high expression of E-cadherin in thyroid cells is reported [28]. In PTC, the low E-cadherin expression is associated with tumor size, multifocality, capsular invasion, extrathyroidal extension, local recurrence and lymph nodes metastases [26, 27, 41, 56–59]. Consequently, the absence of E-cadherin could be regarded as a negative prognostic factor.

However, few papers analyze the E-cadherin profile in different histological subtypes of PTC, with limited results regarding conventional, follicular, tall cell and diffuse sclerosing variant, respectively [27, 36, 58, 60, 61]. These results indicate that the PTC variants associated with poor outcome present lower level of E-cadherin in comparison with the conventional and follicular subtypes, and also with minimally invasive FTC.

To the best of our knowledge, the differences in E-cadherin expression in all histological subtypes of PTC are still not established. Thus, our research brings new data on this topic, based on the particularities of the study group that includes a large variability of histological subtypes of PTC, divided in low- and high-risk groups. Our data show that the E-cadherin expression (low *versus* high) is significantly correlated with the histological risk groups. In our opinion, this assumption sustains the impact of the cellular pattern of PTC on the tumor behavior and the potential prognostic value of the histological variants. Moreover, the present study adds supplementary proofs

that support the relationship between E-cadherin expression and tumor aggressiveness, reflected by tumor size, tumor stage, lymph node metastases, vascular invasion and tumor relapse, in concordance with the previous reported papers.

In TC, β -catenin is less studied than E-cadherin. A limited number of reports show a strong membranous β -catenin pattern in normal thyroid tissue or benign lesions, whereas in PTC, FTC and anaplastic TC the staining is heterogeneously positive in the plasma membrane, cytoplasm or nuclei [34-36, 38]. The main types of TC present a significantly lower membranous β -catenin expression in comparison with the normal or benign thyroid, correlated with the tumor stage, extrathyroidal extension and distant metastasis [34, 35]. In poorly and undifferentiated TC, nuclear β -catenin appears concomitantly with the loss of membranous expression [35, 38, 39]. This fact represents the hallmark for the activation of Wnt/ β -catenin signaling pathway, the nuclear β -catenin operating as a transcriptional activator [33]. Aberrant β -catenin expression or localization are also associated with a worse course in papillary thyroid microcarcinomas [37]. Thus, the changes in β -catenin expression indicate a progressive loss of tumor differentiation with results in a poor prognostic [35, 37–39].

Strictly referring to PTC, β -catenin was analyzed in conventional, follicular, tall cell and diffuse sclerosing variants [36, 62], showing a predominant membranous expression in all subtypes and an infrequent dot-like cytoplasmic (paranuclear) expression in tall cell subtype; the nuclear expression, completely absent in these subtypes [62], is reported only in cribriform-morular variant [39, 63]. The scarcity of reports concerning the β -catenin expression in different histological subtypes of PTC justifies our work directed on this topic, with preliminary results communicated in 2018 at the European Congress of Pathology [40]. The novelty of our study consists in the twofold analysis of β -catenin (membranous- and cytoplasmic-oriented) in different histological variants of PTC, classified as high and low risk, and the comparison of these expression by referring to the clinicopathological factors. Our data reveal significant statistically differences between the membranous β -catenin expression in the two risk groups. Moreover, our results sustain the value

of the membranous β -catenin, assessed as high and low, in relationship with tumor size and tumor stage. Similar results prove, in PTC, the association of low membranous β -catenin pattern with an increased tumor size and distant metastases [34]. The absence of significant statistically differences between the β -catenin cytoplasmic expression and clinicopathological factors could be explained through an early stage of Wnt pathway activation, not necessarily reflected by the cytoplasmic (or nuclear) translocation of β -catenin.

→ Conclusions

Our study demonstrates that E-cadherin and β -catenin expressions differ in low- and high-risk groups of PTC. The aggressive behavior of the high-risk histological variants is associated with reduced membranous E-cadherin, and loss of membranous β -catenin followed by enhanced cytoplasmic expression. These results open large standpoints for a deeper characterization of the histological variants of PTC.

Conflict of interests

The authors declare no conflict of interests.

Acknowledgments

This work was partially funded by "Grigore T. Popa" University of Medicine and Pharmacy under Grant No. 31584/2015.

References

- [1] Lloyd RV, Osamura RY, Klöppel G, Rosai J. World Health Organization (WHO) Classification of Tumours of Endocrine Organs. 4th edition, International Agency for Research on Cancer (IARC) Press, Lyon, 2017, 65–142.
- [2] Kilfoy BA, Zheng T, Holford TR, Han X, Ward MH, Sjodin A, Zhang Y, Bai Y, Zhu C, Guo GL, Rothman N, Zhang Y. International patterns and trends in thyroid cancer incidence, 1973–2002. Cancer Causes Control, 2009, 20(5):525–531.
- [3] LiVolsi VA, Albores-Saavedra J, Asa SL, Baloch ZW, Sobrinho-Simões M, Wenig B, DeLellis RA, Cady B, Mazzaferri EL, Hay I, Fagin JA, Weber AL, Caruso P, Voutilainen PE, Franssila KO, Williams ED, Schneider AB, Nikiforov YE, Rabes HM, Akslen L, Ezzat S, Santoro M, Eng C, Harach HR. Papillary carcinoma. De Lellis RA, Lloyd RV, Heitz PU, Eng C (eds). Pathology and genetics of tumours of endocrine organs. WHO Classification of Tumours, IARC Press, Lyon, 2004, 57–66.
- [4] Sethi K, Sarkar S, Das S, Mohanty B, Mandal M. Biomarkers for the diagnosis of thyroid cancer. J Exp Ther Oncol, 2010, 8(4):341–352.
- [5] Nikiforov YE. Molecular analysis of thyroid tumors. Mod Pathol, 2011, 24(Suppl 2):S34–S43.
- [6] Radu TG, Mogoantă L, Busuioc CJ, Stănescu C, Grosu F. Histological and immunohistochemical aspects of papillary thyroid cancer. Rom J Morphol Embryol, 2015, 56(2 Suppl): 789–795.
- [7] Radu TG, Ciurea ME, Mogoantă SŞ, Busuioc CJ, Grosu F, Ţenovici M, Petrescu IO, Vladu IM. Papillary thyroid cancer stroma – histological and immunohistochemical study. Rom J Morphol Embryol, 2016, 57(2 Suppl):801–809.
- [8] De Lellis R, Williams E. Thyroid and parathyroid tumours. In: De Lellis RA, Lloyd RV, Heitz PU, Eng C (eds). Pathology and genetics of tumours of endocrine organs. WHO Classification of Tumours, IARC Press, Lyon, 2004, 50–66.
- [9] Dean DS, Hay ID. Prognostic indicators in differentiated thyroid carcinoma. Cancer Control, 2000, 7(3):229–239.
- [10] Sherman SI. Thyroid carcinoma. Lancet, 2003, 361(9356): 501–511.
- [11] Kakudo K, Tang W, Ito Y, Mori I, Nakamura Y, Miyauchi A. Papillary carcinoma of the thyroid in Japan: subclassification of common type and identification of low risk group. J Clin Pathol, 2004, 57(10):1041–1046.

- [12] LiVolsi VA. Papillary thyroid carcinoma: an update. Mod Pathol, 2011, 24(Suppl 2):S1–S9.
- [13] Lloyd RV, Buehler D, Khanafshar E. Papillary thyroid carcinoma variants. Head Neck Pathol, 2011, 5(1):51–56.
- [14] Kakudo K, Wakasa T, Ohta Y, Yane K, Satoh S, Yokoi T, Yamashita H. Moderate-risk and high-risk thyroid carcinomas of follicular cell origin. J Basic Clin Med, 2014, 3(1):12–17.
- [15] Pećina-Slaus N. Tumor suppressor gene E-cadherin and its role in normal and malignant cells. Cancer Cell Int, 2003, 3(1):17.
- [16] Ozawa M, Baribault H, Kemler R. The cytoplasmic domain of the cell adhesion molecule uvomorulin associates with three independent proteins structurally related in different species. EMBO J, 1989, 8(6):1711–1717.
- [17] McCrea PD, Turck CW, Gumbiner B. A homolog of the armadillo protein in *Drosophila* (plakoglobin) associated with E-cadherin. Science, 1991, 254(5036):1359–1361.
- [18] Lilien J, Balsamo J. The regulation of cadherin-mediated adhesion by tyrosine phosphorylation/dephosphorylation of beta-catenin. Curr Opin Cell Biol, 2005, 17(5):459–465.
- [19] Brembeck FH, Rosário M, Birchmeier W. Balancing cell adhesion and Wnt signaling, the key role of β-catenin. Curr Opin Genet Dev, 2006, 16(1):51–59.
- [20] Bracke ME, Van Roy FM, Mareel MM. The E-cadherin/catenin complex in invasion and metastasis. Curr Top Microbiol Immunol, 1996, 213(Pt 1):123–161.
- [21] Kefeli M, Karagöz F, Barış S, Yıldız L, Aydın O, Kandemir B. [E-cadherin and Ki-67 expression in colorectal carcinomas and their relationship with stage, histological subtype and grade]. Turk J Pathol, 2005, 21(1–2):8–10.
- [22] Hirohashi S, Kanai Y. Cell adhesion system and human cancer morphogenesis. Cancer Sci, 2003, 94(7):575–581.
- [23] Guilford P. E-cadherin downregulation in cancer: fuel on the fire? Mol Med Today, 1999, 5(4):172–177.
 [24] Berx G, Van Roy F. The E-cadherin/catenin complex: an
- [24] Berx G, Van Roy F. The E-cadherin/catenin complex: an important gatekeeper in breast cancer tumorigenesis and malignant progression. Breast Cancer Res, 2001, 3(5):289– 293
- [25] Sobrinho-Simões M, Oliveira C. Different types of epithelial cadherin alterations play different roles in human carcinogenesis. Adv Anat Pathol, 2002, 9(6):329–337.
- [26] Rocha AS, Soares P, Fonseca E, Cameselle-Teijeiro J, Oliveira MC, Sobrinho-Simões M. E-cadherin loss rather than beta-catenin alterations is a common feature of poorly differentiated thyroid carcinomas. Histopathology, 2003, 42(6): 580–587.
- [27] Ceyran AB, Şenol S, Şimşek BÇ, Sağıroğlu J, Aydın A. Role of CD56 and E-cadherin expression in the differential diagnosis of papillary thyroid carcinoma and suspected follicular-patterned lesions of the thyroid: the prognostic importance of E-cadherin. Int J Clin Exp Pathol, 2015, 8(4): 3670–3680.
- [28] Liu H, Lin F. Application of immunohistochemistry in thyroid pathology. Arch Pathol Lab Med, 2015, 139(1):67–82.
- [29] Kawasaki Y, Sato R, Akiyama T. Mutated APC and Asef are involved in the migration of colorectal tumour cells. Nat Cell Biol, 2003, 5(3):211–215.
- [30] Khramtsov AI, Khramtsova GF, Tretiakova M, Huo D, Olopade OI, Goss KH. Wnt/beta-catenin pathway activation is enriched in basal-like breast cancers and predicts poor outcome. Am J Pathol, 2010, 176(6):2911–2920.
- [31] Sinnberg T, Menzel M, Ewerth D, Sauer B, Schwarz M, Schaller M, Garbe C, Schittek B. β-Catenin signaling increases during melanoma progression and promotes tumor cell survival and chemoresistance. PLoS One, 2011, 6(8):e23429.
- [32] Tao J, Calvisi DF, Ranganathan S, Cigliano A, Zhou L, Singh S, Jiang L, Fan B, Terracciano L, Armeanu-Ebinger S, Ribback S, Dombrowski F, Evert M, Chen X, Monga SPS. Activation of β-catenin and Yap1 in human hepatoblastoma and induction of hepatocarcinogenesis in mice. Gastroenterology, 2014, 147(3):690–701.
- [33] Shang S, Hua F, Hu ZW. The regulation of β-catenin activity and function in cancer: therapeutic opportunities. Oncotarget, 2017, 8(20):33972–33989.
- [34] Böhm J, Niskanen L, Kiraly K, Kellokoski J, Eskelinen M, Hollmen S, Alhava E, Kosma VM. Expression and prognostic value of alpha-, beta-, and gamma-catenins in differentiated thyroid carcinoma. J Clin Endocrinol Metab, 2000, 85(12): 4806–4811.

- [35] Garcia-Rostan G, Camp RL, Herrero A, Carcangiu ML, Rimm DL, Tallini G. β-Catenin dysregulation in thyroid neoplasms: downregulation, aberrant nuclear expression, and CTNNB1 exon 3 mutations are markers for aggressive tumor phenotypes and poor prognosis. Am J Pathol, 2001, 158(3):987–996.
- [36] Rocha AS, Soares P, Seruca R, Máximo V, Matias-Guiu X, Cameselle-Teijeiro J, Sobrinho-Simões M. Abnormalities of the E-cadherin/catenin adhesion complex in classical papillary thyroid cancer and in its diffuse sclerosing variant. J Pathol, 2001, 194(3):358–366.
- [37] Lantsov D, Meirmanov S, Nakashima M, Kondo H, Saenko V, Naruke Y, Namba H, Ito M, Abrosimov A, Lushnikov E, Sekine I, Yamashita Sh. Cyclin D1 overexpression in thyroid papillary microcarcinoma: its association with tumour size and aberrant beta-catenin expression. Histopathology, 2005, 47(3):248–256.
- [38] Ralhan R, Cao J, Lim T, MacMillan C, Freeman JL, Walfish PG. EpCAM nuclear localization identifies aggressive thyroid cancer and is a marker for poor prognosis. BMC Cancer, 2010. 10:331.
- [39] Lam AK, Saremi N. Cribriform-morular variant of papillary thyroid carcinoma: a distinctive type of thyroid cancer. Endocr Relat Cancer, 2017, 24(4):R109–R121.
- [40] Caruntu ID, Giusca SE, Lozneanu L, Apostol Ciobanu D, Andriescu EC. Expression of β-catenin in the papillary thyroid carcinoma. Virchow Archiv, 2018, 473(1 Suppl):S252–S253.
- [41] Wiseman SM, Masoudi H, Niblock P, Turbin D, Rajput A, Hay J, Filipenko D, Huntsman D, Gilks B. Derangement of the E-cadherin/catenin complex is involved in transformation of differentiated to anaplastic thyroid carcinoma. Am J Surg, 2006, 191(5):581–587.
- [42] Ciobanu Apostol D, Căruntu ID, Lozneanu L, Andriescu EC, Giuşcă SE. HER-2/neu expression in different histological subtypes of papillary thyroid carcinoma. Rom J Morphol Embryol, 2017, 58(2):439–444.
- [43] Giusca SE, Amalinei C, Lozneanu L, Ciobanu Apostol D, Andriescu EC, Scripcariu A, Balan R, Avadanei ER, Căruntu ID. Heterogeneous periostin expression in different histological variants of papillary thyroid carcinoma. BioMed Res Int, 2017, 2017:8701386.
- [44] Ciobanu D, Căruntu ID, Lozneanu L, Andriescu EC, Leuştean L, Giuşcă SE. Tubulin, a possible marker for the prognostic stratification and therapy in papillary thyroid carcinoma. Farmacia, 2018, 66(4):635–643.
- [45] Perou CM, Sørlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, Fluge O, Pergamenschikov A, Williams C, Zhu SX, Lønning PE, Børresen-Dale AL, Brown PO, Botstein D. Molecular portraits of human breast tumours. Nature, 2000, 406(6797):747–752.
- [46] West L, Vidwans SJ, Campbell NP, Shrager J, Simon GR, Bueno R, Dennis PA, Otterson GA, Salgia R. A novel classification of lung cancer into molecular subtypes. PLoS One, 2012, 7(2):e31906.
- [47] Heuberger J, Birchmeier W. Interplay of cadherin-mediated cell adhesion and canonical Wnt signaling. Cold Spring Harb Perspect Biol, 2010, 2(2):a002915.
- [48] Brabant G, Hoang-Vu C, Cetin Y, Dralle H, Scheumann G, Mölne J, Hansson G, Jansson S, Ericson LE, Nilsson M. E-cadherin: a differentiation marker in thyroid malignancies. Cancer Res, 1993, 53(20):4987–4993.
- [49] Soares P, Berx G, van Roy F, Sobrinho-Simões M. E-cadherin gene alterations are rare events in thyroid tumors. Int J Cancer, 1997, 70(1):32–38.

- [50] von Wasielewski R, Rhein A, Werner M, Scheumann GFW, Dralle H, Pötter E, Brabant G, Georgii A. Immunohistochemical detection of E-cadherin in differentiated thyroid carcinomas correlates with clinical outcome. Cancer Res, 1997, 57(12):2501–2507.
- [51] Walgenbach S, Sternheim E, Bittinger F, Görges R, Andreas J, Junginger T. [Prognostic value of E-cadherin in papillary thyroid carcinoma]. Chirurg, 1998, 69(2):186–190.
- [52] Naito A, Iwase H, Kuzushima T, Nakamura T, Kobayashi S. Clinical significance of E-cadherin expression in thyroid neoplasms. J Surg Oncol, 2001, 76(3):176–180.
- [53] Brecelj E, Frković Grazio S, Auersperg M, Bračko M. Prognostic value of E-cadherin expression in thyroid follicular carcinoma. Eur J Surg Oncol, 2005, 31(5):544–548.
- [54] Choi YL, Kim MK, Suh JW, Han J, Kim JH, Yang JH, Nam SJ. Immunoexpression of HBME-1, high molecular weight cytokeratin, cytokeratin 19, thyroid transcription factor-1, and E-cadherin in thyroid cancers. J Korean Med Sci, 2005, 20(5): 853–859
- [55] Kato N, Tsuchiya T, Tamura G, Motoyama T. E-cadherin expression in follicular carcinoma of the thyroid. Pathol Int, 2002. 52(1):13–18.
- [56] Erdem H, Gündogdu C, Sipal S. Correlation of E-cadherin, VEGF, COX-2 expression to prognostic parameters in papillary thyroid carcinoma. Exp Mol Pathol, 2011, 90(3):312–317.
- [57] Sethi K, Sarkar S, Das S, Rajput S, Mazumder A, Roy B, Patra S, Mohanty B, El-Naggar AK, Mandal M. Expressions of CK-19, NF-kappaB, E-cadherin, beta-catenin and EGFR as diagnostic and prognostic markers by immunohistochemical analysis in thyroid carcinoma. J Exp Ther Oncol, 2011, 9(3): 187–199.
- [58] Calangiu CM, Simionescu CE, Stepan AE, Cernea D, Zăvoi RE, Mărgăritescu C. The expression of CK19, vimentin and E-cadherin in differentiated thyroid carcinomas Rom J Morphol Embryol, 2014, 55(3):919–925.
- [59] Ivanova K, Ananiev J, Aleksandrova E, Ignatova MM, Gulubova M. Expression of E-cadherin/beta-catenin in epithelial carcinomas of the thyroid gland. Open Access Maced J Med Sci, 2017, 5(2):155–159.
- [60] Ito Y, Hirokawa M, Fukushima M, Inoue H, Yabuta T, Uruno T, Kihara M, Higashiyama T, Takamura Y, Miya A, Kobayashi K, Matsuzuka F, Miyauchi A. Prevalence and prognostic significance of poor differentiation and tall cell variant in papillary carcinoma in Japan. World J Surg, 2008, 32(7):1535–1543; discussion 1544–1545.
- [61] Slowinska-Klencka D, Sporny S, Stasikowska-Kanicka O, Popowicz B, Klencki M. E-cadherin expression is more associated with histopathological type of thyroid cancer than with the metastatic potential of tumors. Folia Histochem Cytobiol, 2012, 50(4):519–526.
- [62] Min HS, Lee C, Jung KC. Correlation of immunohistochemical markers and BRAF mutation status with histological variants of papillary thyroid carcinoma in the Korean population. J Korean Med Sci, 2013, 28(4):534–541.
- [63] Jung CK, Choi YJ, Lee KY, Bae JS, Kim HJ, Yoon SK, Son YI, Chung JH, Oh YL. The cytological, clinical, and pathological features of the cribriform-morular variant of papillary thyroid carcinoma and mutation analysis of CTNNB1 and BRAF genes. Thyroid, 2009, 19(8):905–913.

Corresponding author

Irina-Draga Căruntu, Professor, MD, PhD, Department of Morphofunctional Sciences I, "Grigore T. Popa" University of Medicine and Pharmacy, 16 University Street, 700115 Iaşi, Romania; Phone +40727–003 700, e-mails: irina.caruntu@umfiasi.ro, irinadragacaruntu@gmail.com

Received: January 30, 2018 Accepted: November 26, 2018